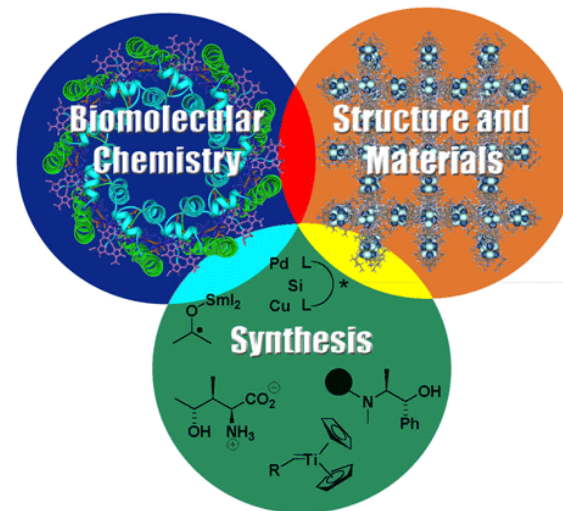




University of Glasgow



Chemical Biology: Oxidative Processes of Ageing and the Potential of Antioxidants



37th Annual Meeting of American Aging Association
Boulder, Colorado, 31st May 2008

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West
CHEM

Chronological and Biological ageing do not always equate.....



57



43



61




52

The Root of the Problem

- Oxidative Stress leads to Damage
- Damage decreases ability to deal with further stress
- Responsible for cellular ageing
- Causes or exacerbates diseases of old age
 - neurodegeneration
 - stroke
 - heart disease
 - diabetes

Reactive Oxygen Species

<u>Couple (Oxidant / Antioxidant)</u>	<u>E° / mV</u>	
HO \cdot , H $^{+}$ / H $_2$ O	2310	<div style="display: flex; align-items: center; justify-content: center;"> <div style="margin-right: 10px;">Highly oxidising</div> <div style="text-align: center;">  </div> </div>
RO \cdot , H $^{+}$ / ROH	1600	
HOO \cdot , H $^{+}$ / H $_2$ O $_2$	1150	
ROO \cdot , H $^{+}$ (lipid peroxy radical) / ROOH	1060	
O $_2^{\cdot-}$, 2H $^{+}$ (superoxide anion) / H $_2$ O $_2$	940	
PUFA \cdot , H $^{+}$ / PUFA-H (polyunsaturated fatty acid)	600	
α -tocopheroxyl \cdot , H $^{+}$ / α -tocopherol (Vit E)	500	
Ascorbate \cdot , H $^{+}$ / ascorbate		
monoanoin (Vit C)	282	

Buttner, G. *Arch. Biochem. Biophys.*, **1993**, 535

Cellular Ageing (Senescence):

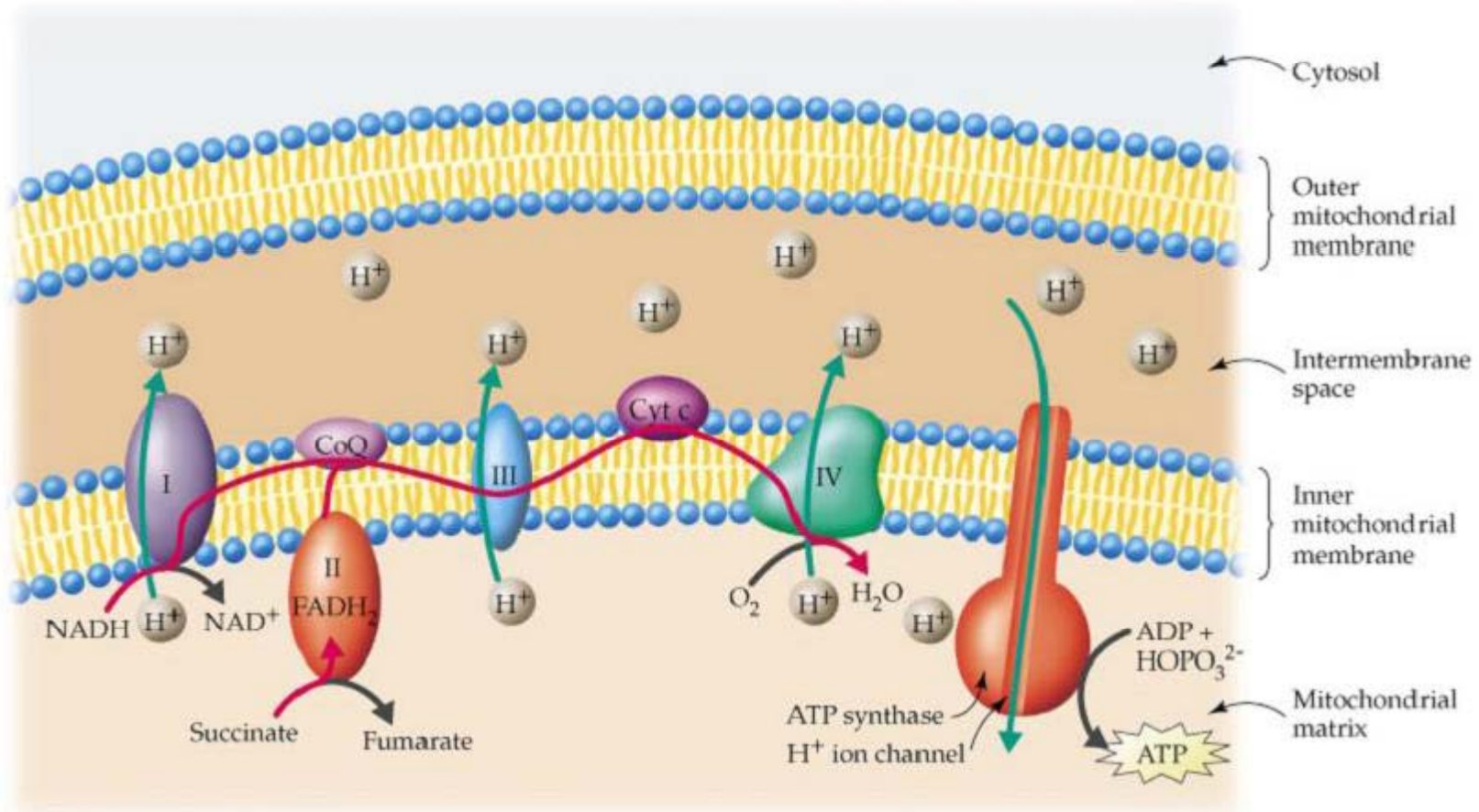
- Damage to genes and the biological clock (**telomeres**)
- Damage to membranes
- Damage to metabolic capacity - **oxidative damage to mitochondria**



Reactive Oxygen Species

– produced in mitochondria

– damage mitochondria including mitochondrial DNA



Chemical Biology

Detect and identify the contributors to oxidative stress

- EPR spectroscopy - the only technique that sees radicals directly

Locate and quantify the oxidizing species

- Probes targeted to Mitochondria

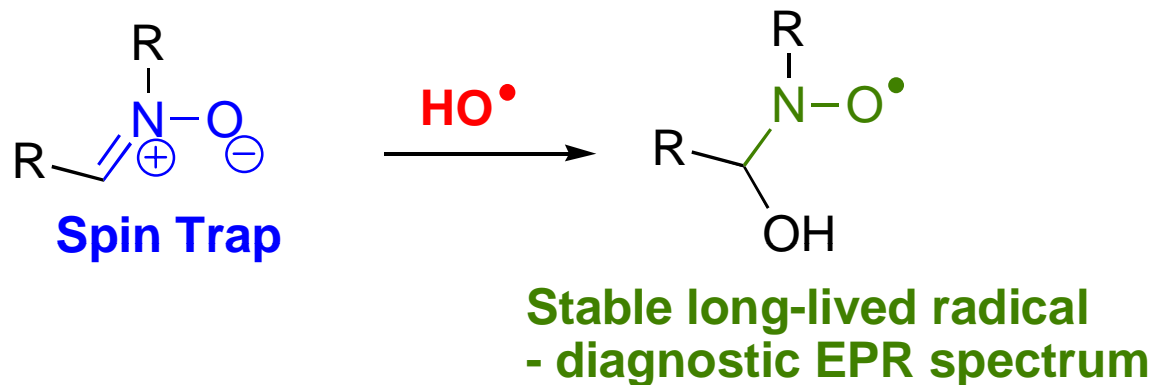
Ameliorate oxidative stress

- therapeutic antioxidants that scavenge ROS:

Track compounds claimed to be good for you

- Assess whether antioxidants found in food can be biological antioxidants *in vivo*.

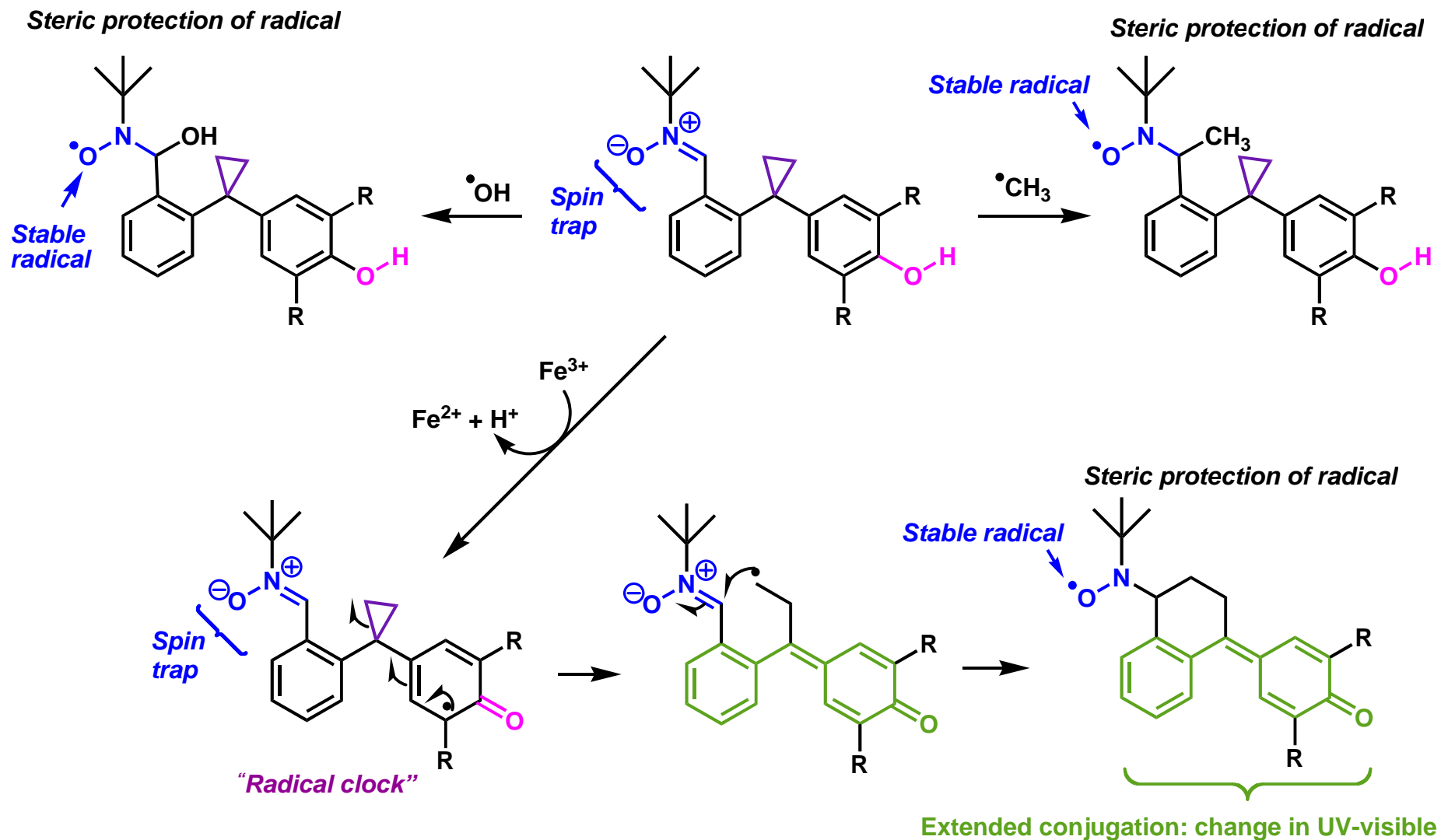
Detect and Identify ROS: Spin traps and EPR Spectroscopy.



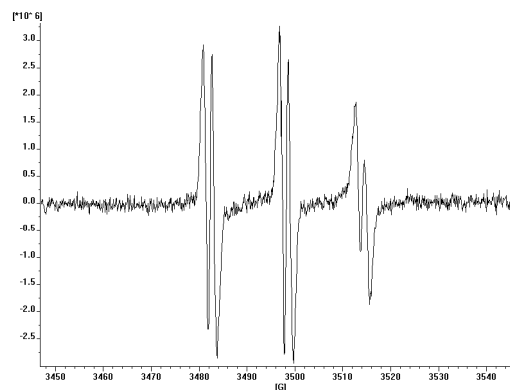
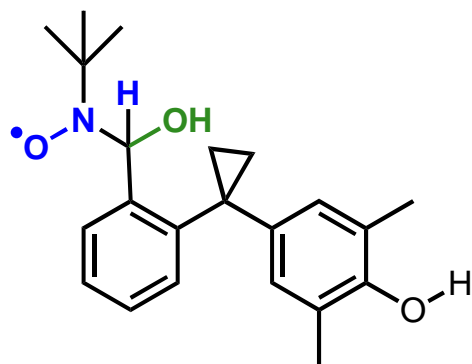
*Detects ROS:
In vitro and in vivo
Only sees the radicals*

How can different contributors to oxidative stress be detected selectively?

Differential Radical Detection and Double Display



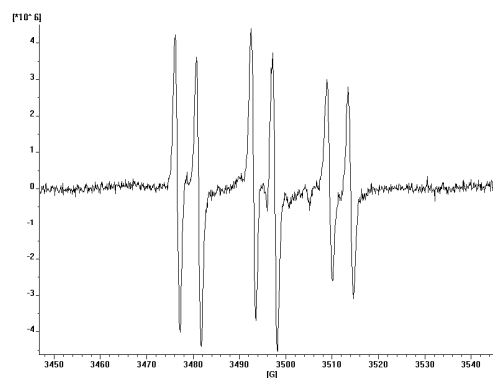
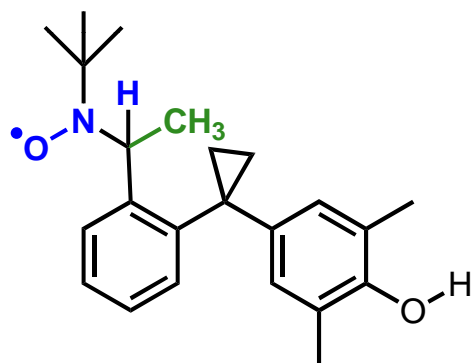
Trapping carbon-centred and oxygen-centred radicals



$$g = 2.0059$$

$$A_N = 15.93 \text{ G (t)}$$

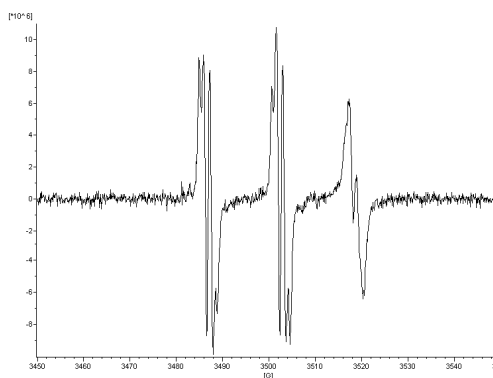
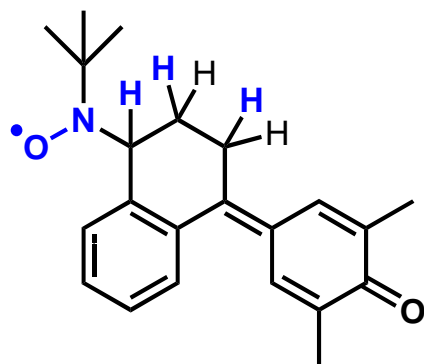
$$A_{H^\beta} = 1.85 \text{ G (d)}$$



$$g = 2.0056$$

$$A_N = 16.32 \text{ G (t)}$$

$$A_{H^\beta} = 4.57 \text{ G (d)}$$



$$g = 2.0055$$

$$A_N = 15.70 \text{ G (t)}$$

$$A_{H^\beta} = 1.42 \text{ G (d)}$$

$$A_{H^\gamma/\delta} = 1.05 \text{ and } 0.99 \text{ G (dd)}$$

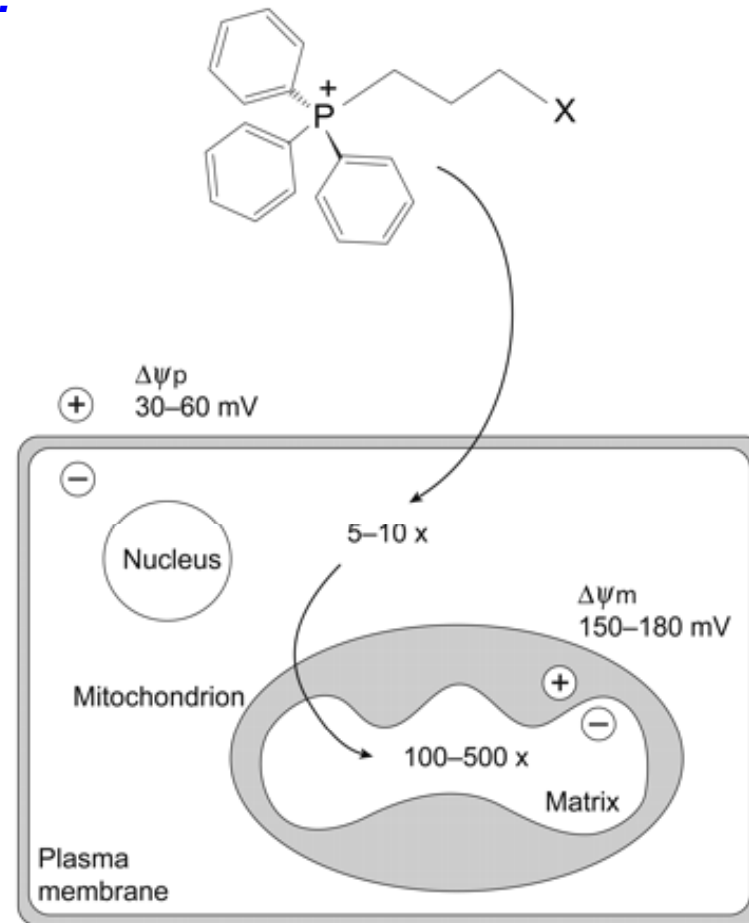
Locate and quantify ROS:

Target Mitochondria

- *Main source of ROS*
- *Mitochondrial dysfunction involved in*
 - *neurodegenerative diseases*
 - *ageing.*

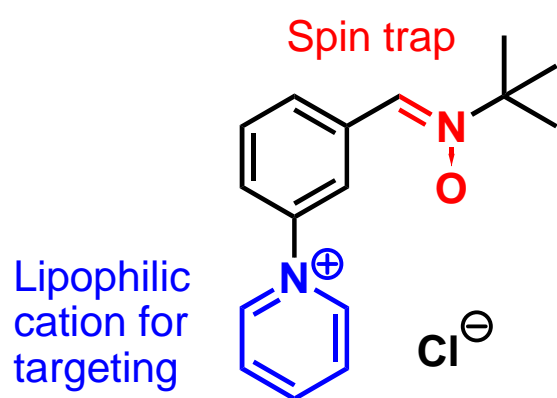
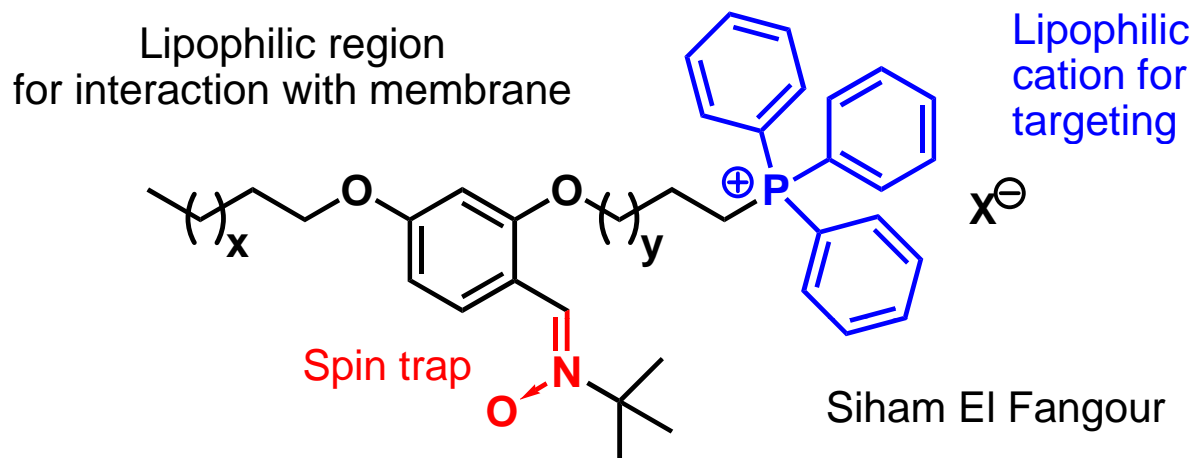
See: Lin, M. T.; Beal, M. F. *Nature* **2006**, *443*, 787-795

Use membrane potential:

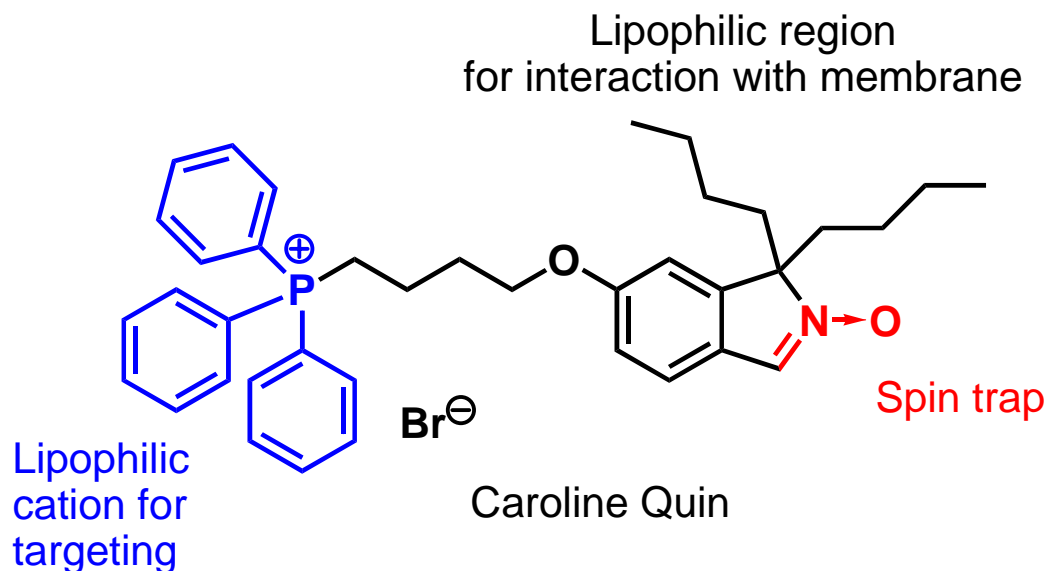


- (a) pH within the matrix about 1.4 units lower than in the cytosol
- (b) Large membrane potential ($\Delta\psi$) of -150 to -180 mV (negative inside the matrix) across the inner membrane of mitochondrion.
- (c) Both contribute to Δp so that this is about 230 mV in active mitochondria.

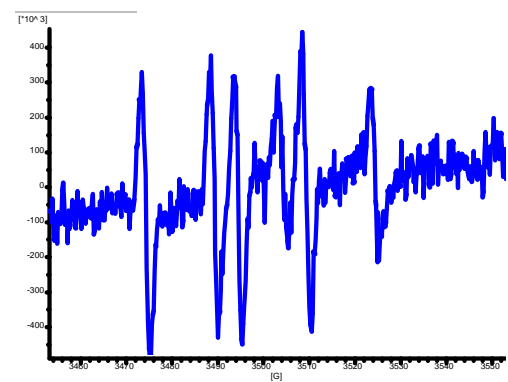
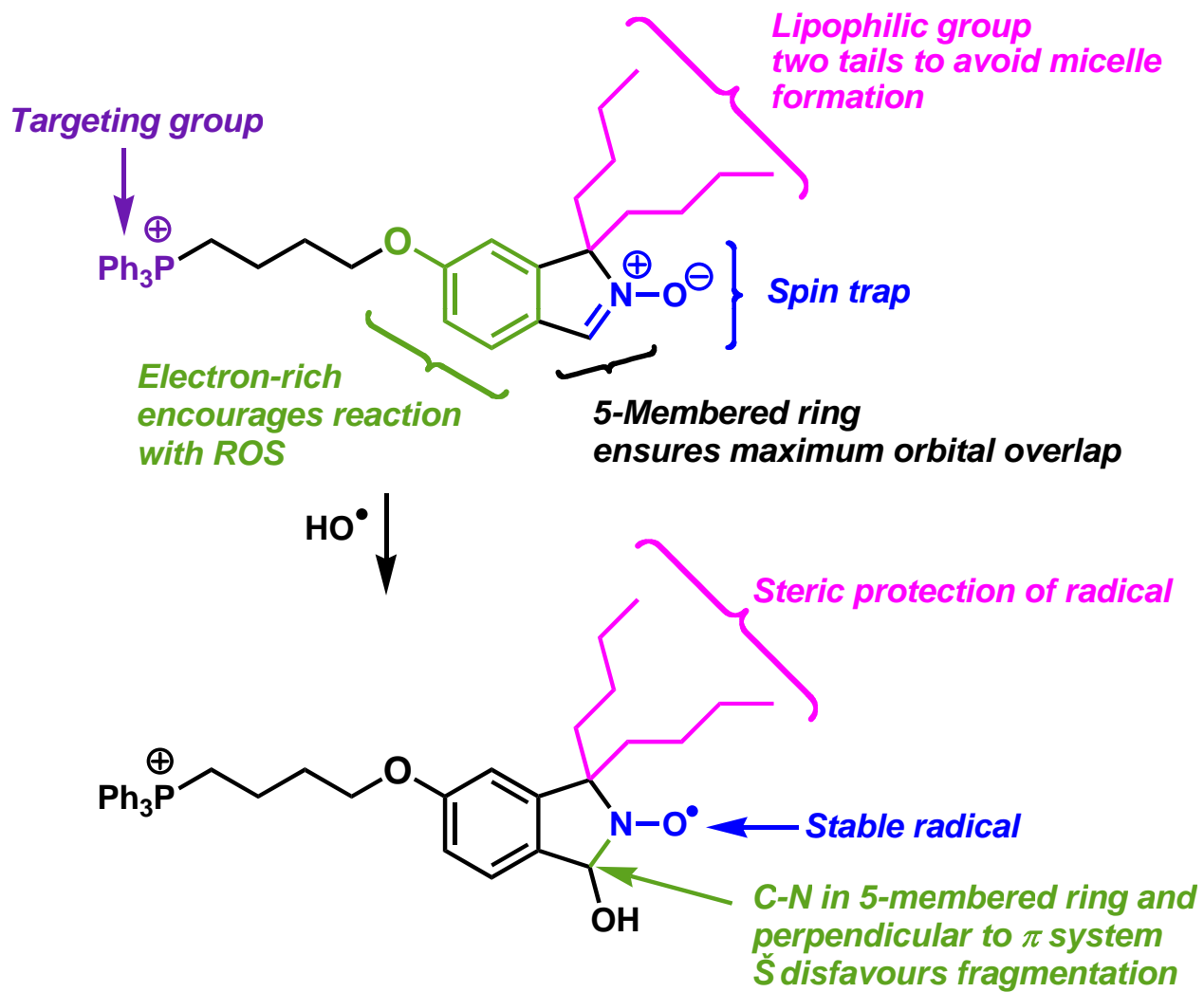
Lipophilic cations: radical detectors/antioxidants targeted to mitochondria



Linsey Robertson



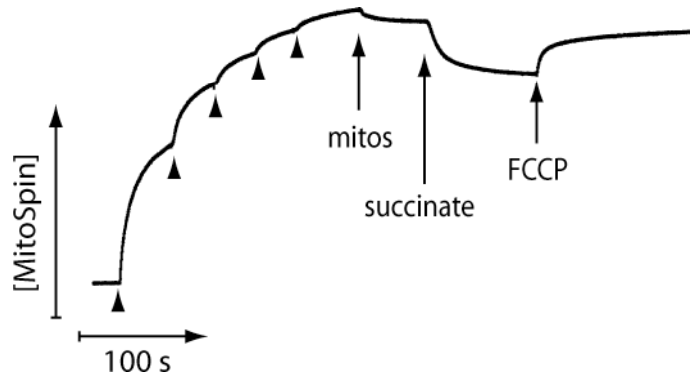
MitoSPIN:



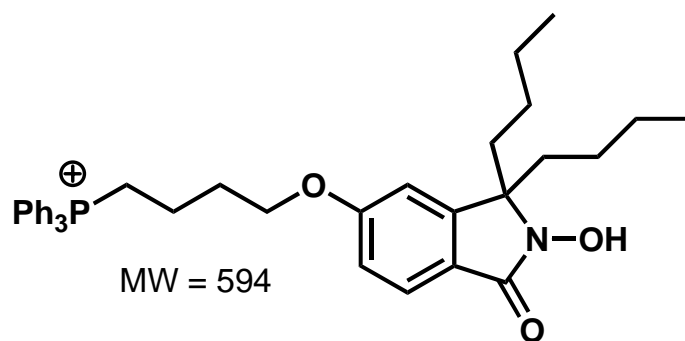
EPR spectrum

MitoSPIN

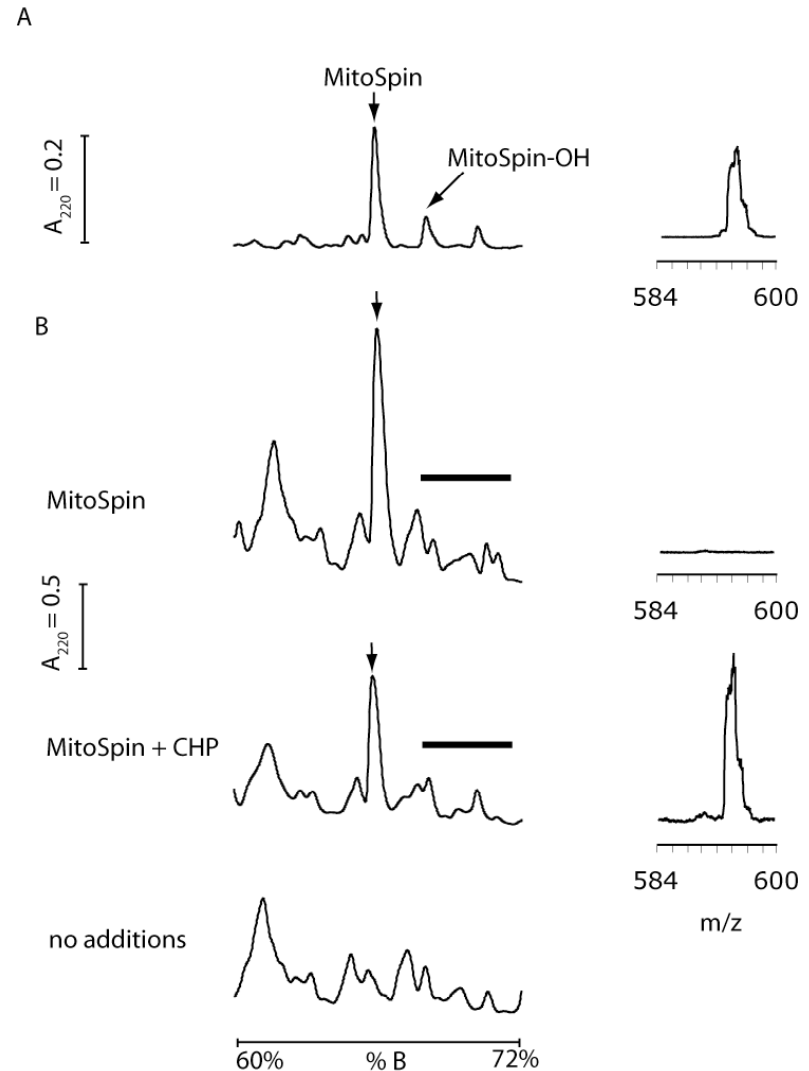
Accumulates 1000-fold in active mitochondria



Is oxidized to MitoSPIN-OH by hydroxyl radicals



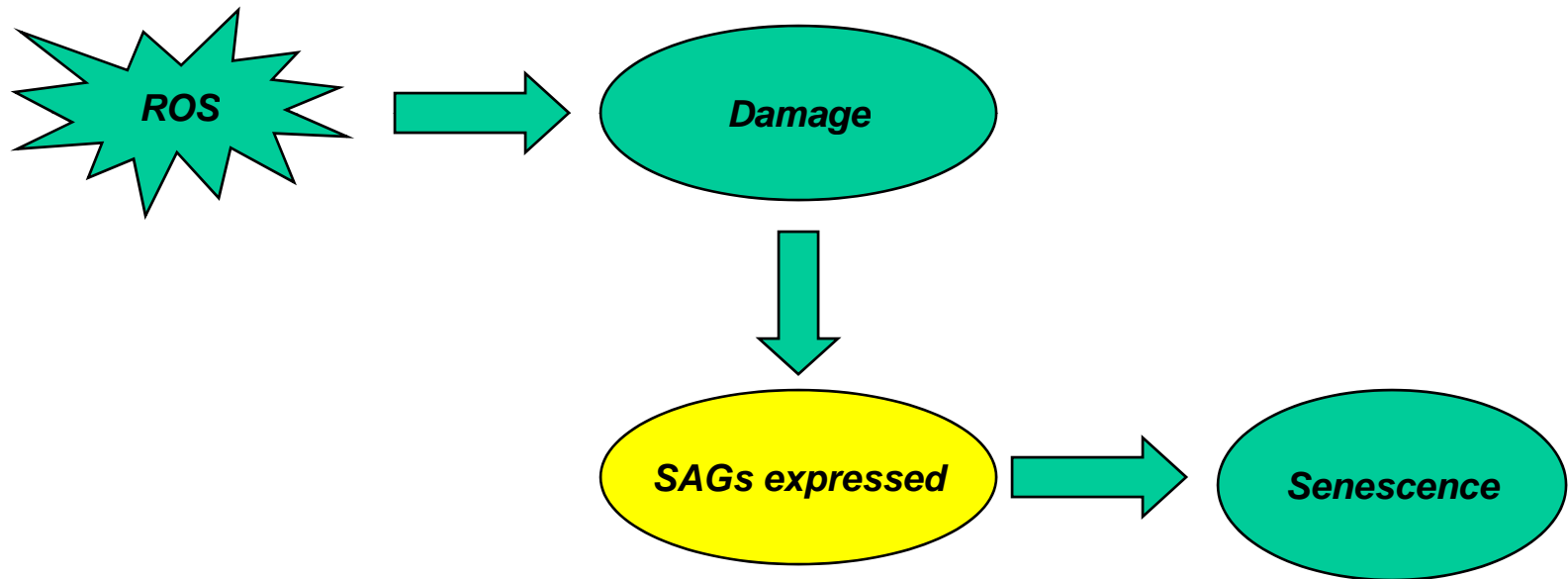
Can be used to measure ROS in mitochondria by ESI-MS



Michael Murphy, Jan Trnka, MRC Human Nutrition Unit, Cambridge

Ameliorate oxidative stress and slow ROS-associated senescence

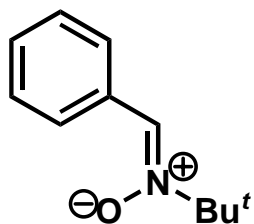
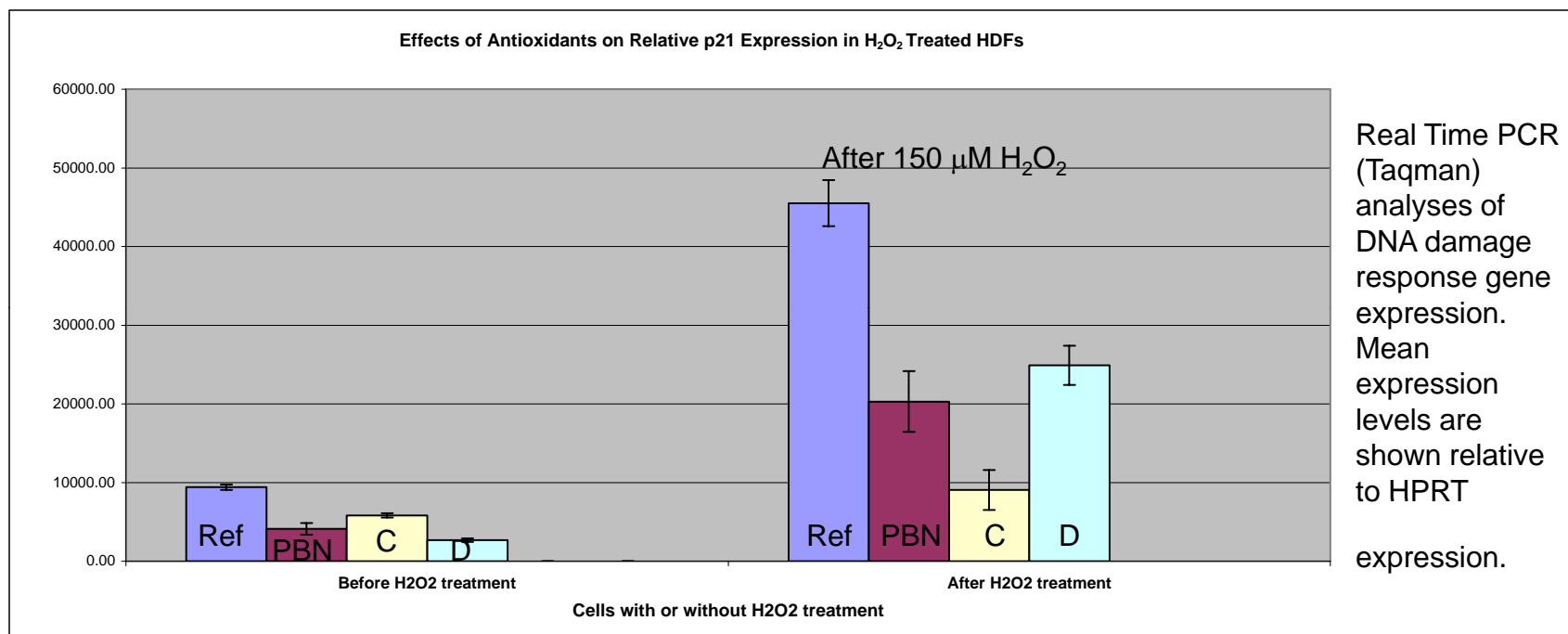
- Oxidative stress increases expression of **s**enescence **a**ssociated **g**enes



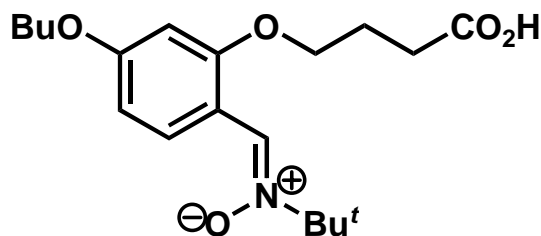
Antioxidants protect primary **H**uman **D**iploid **f**ibroblasts from an oxidative assault?

p21 expression in HDF cells

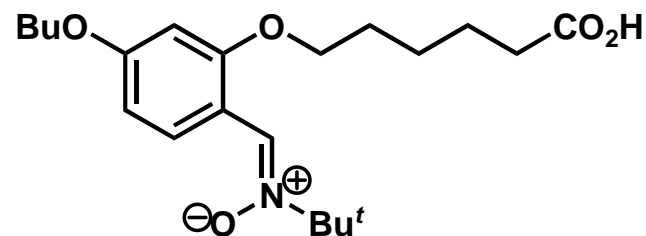
p21 inhibits cyclin dependent kinases in response to p53 and thus initiates senescence



PBN (@ 750 μM)



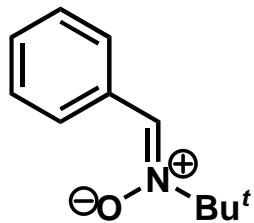
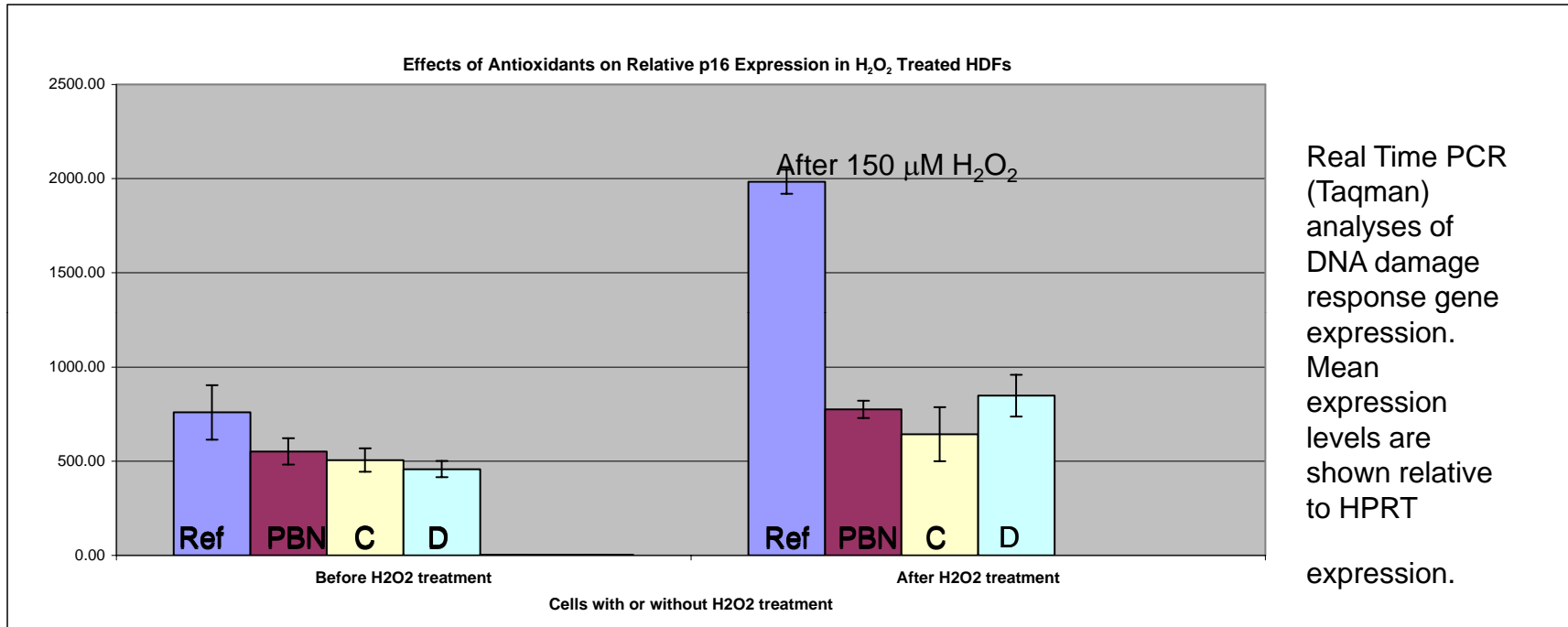
Nitrone C (@ 400 μM)



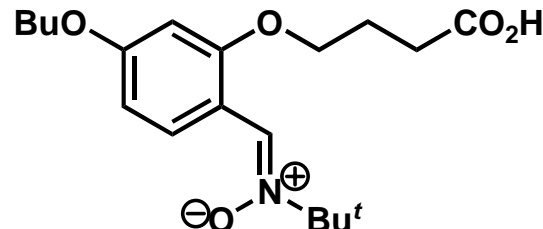
Nitrone D (@ 300 μM)

p16 expression in HDF cells

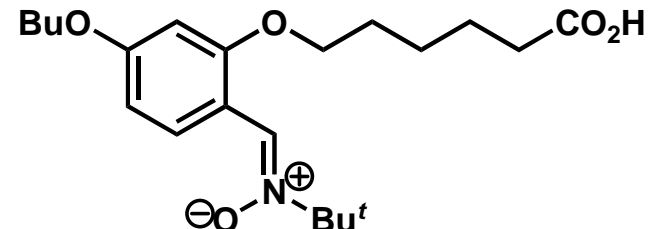
Involved in long term maintenance of senescence



PBN (@ 750 μM)



Nitrone C (@ 400 μM)



Nitrone D (@ 300 μM)

Sklavounou, E.; Hay, A.; Lamb, K.; Brown, E.; MacIntyre, A.; George, W. D.; Hartley R. C.; Shiels, P. G.
BBRC **2006**, 347, 420-427.

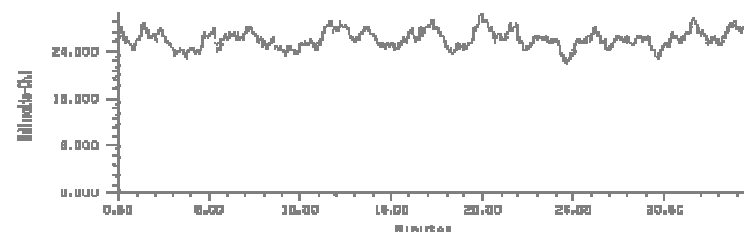
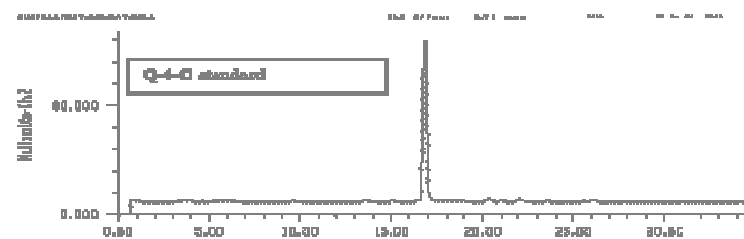
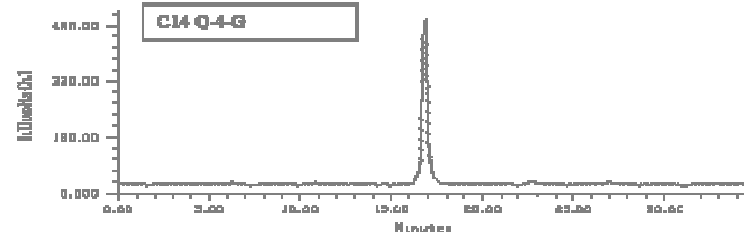
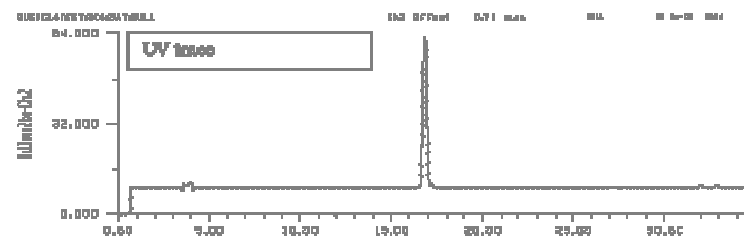
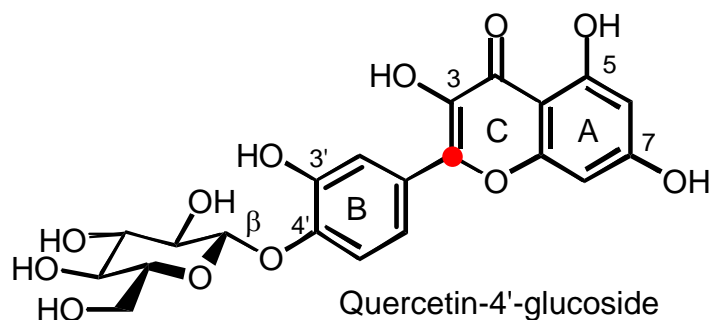
***Tracking dietary compounds
and metabolites***

– isotopic labelling

Are flavonoids biological antioxidants?



Isotopic labelling to determine uptake, distribution and metabolism of drugs and natural products



W. Mullen, R. C. Hartley, A. Crozier, *Journal of Chromatography A* **2003**, *1007*, 21-29. W. Mullen, B. A. Graf, S. T. Caldwell, R. C. Hartley, G. G. Duthie, C. A. Edwards, M. E. J. Lean, and A. Crozier, *J. Agric. Food Chem.* **2002**, *50*, 6902-6909. S. T. Caldwell, A. Crozier, and R. C. Hartley, *Tetrahedron* **2000**, *56*, 4101-4106.

Distribution of Radioactivity in Rats 60 min after Ingestion of [2- ¹⁴C]Quercetin-4'-Glucoside.

Tissue/fluid	Total radioactivity	Concentration of radioactivity
Intestine and contents	54,530 ± 4,728 (93.6%)	2,117 ± 162
Plasma	1,644 ± 157 (2.8%)	110 ± 10
Red blood cells	4 ± 4 (-)	0 ± 0
Liver	684 ± 240 (1.2%)	37 ± 313
Kidneys	468 ± 295 (0.8%)	147 ± 109
Spleen	6 ± 1 (-)	7 ± 1
Brain	1 ± 1 (-)	0 ± 0
Lungs	38 ± 10 (0.07%)	21 ± 5
Heart	18 ± 2 (0.03%)	14 ± 2
Muscle	839 ± 310 (1.4%)	4 ± 2
Testes	25 ± 34 (0.04%)	8 ± 2

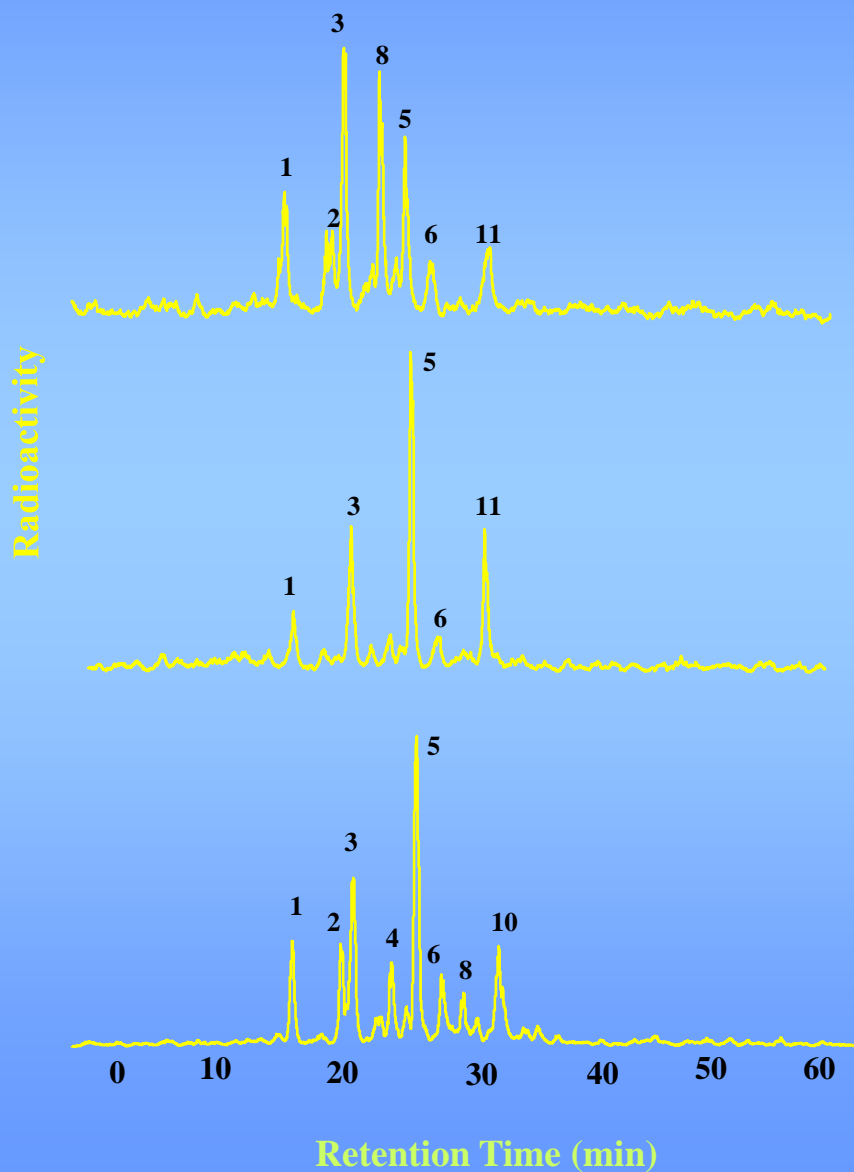
Radioactivity expressed as dpm x 10³ per tissue/fluid per rat ± standard deviation (n = 3) and in parenthesis as a percentage of the recovered radioactivity.

3 mg of quercetin glucoside per 430 g rat (58.6 x 10⁶ dpm)

METABOLISM OF [2-¹⁴C]QUERCETIN-4'-GLUCOSIDE IN RATS

Aqueous extracts

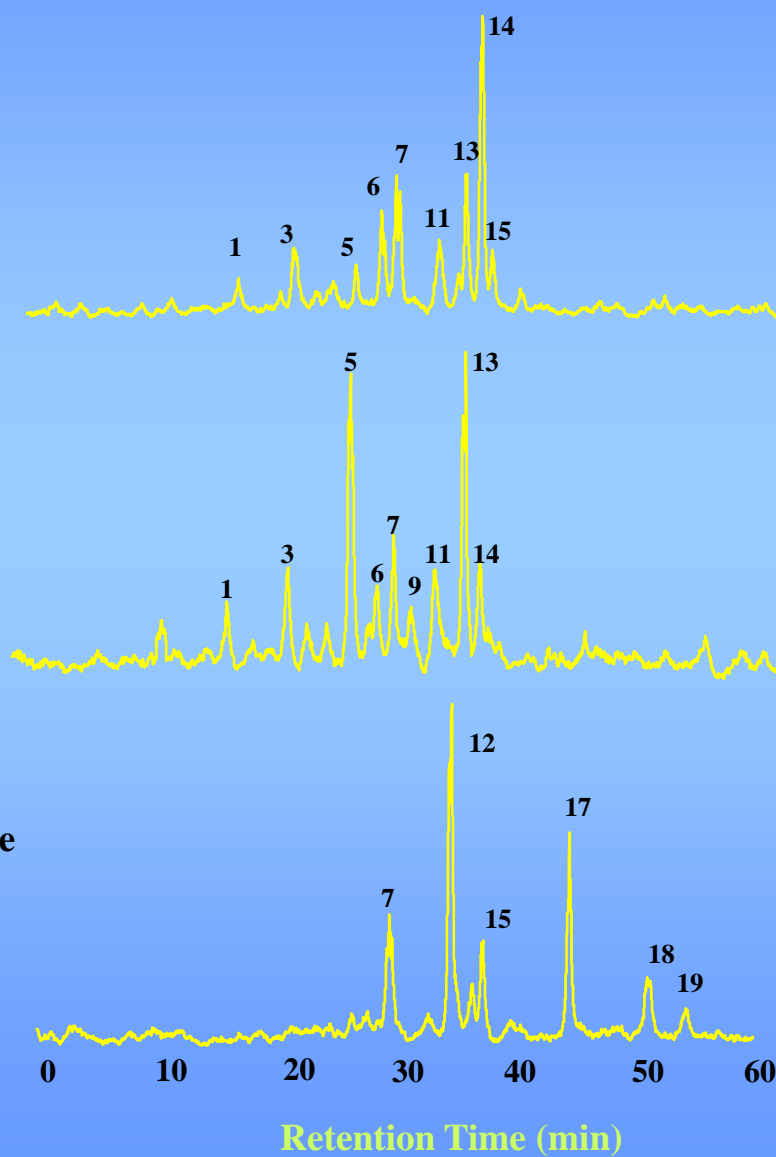
Organic extracts



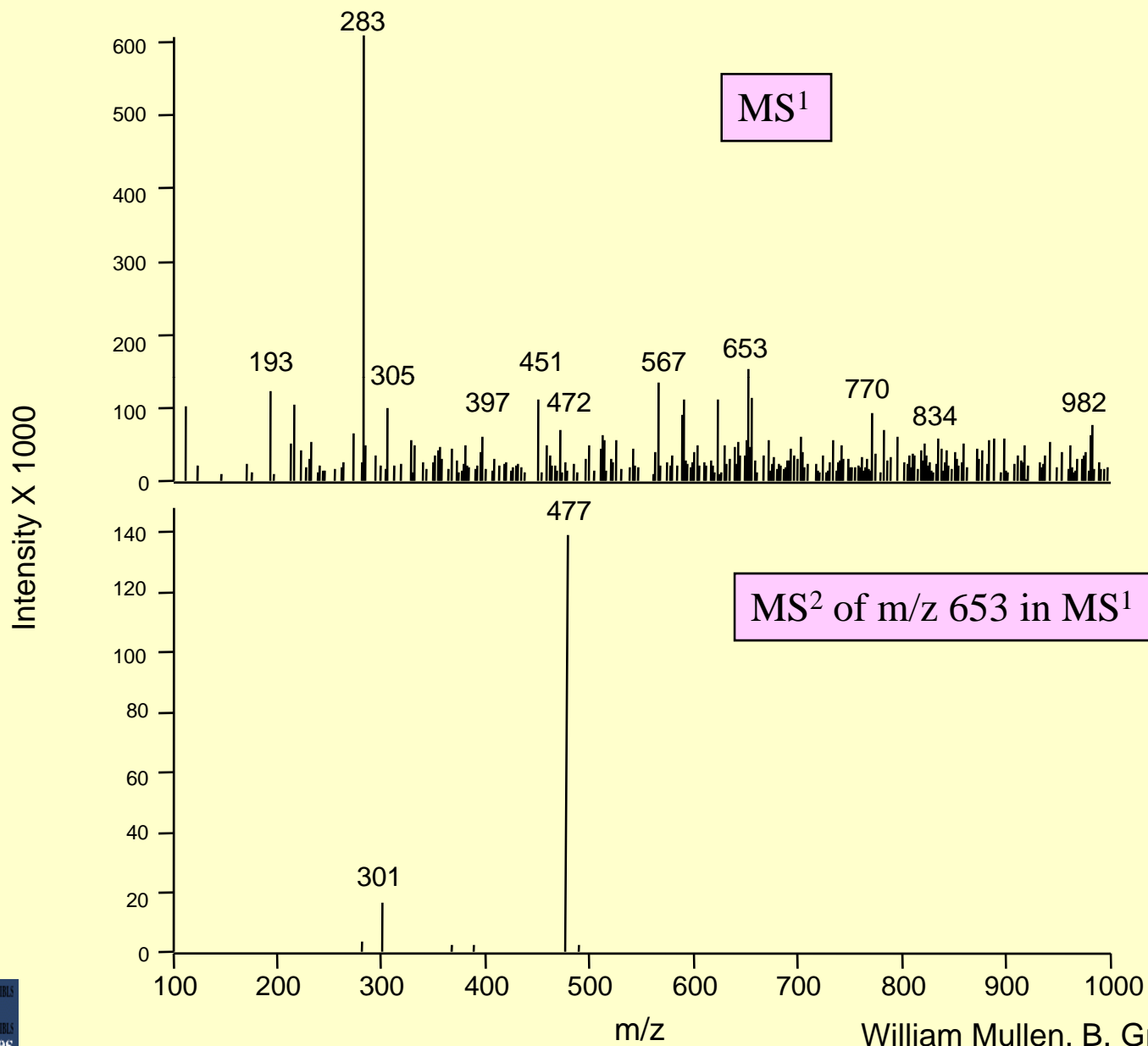
Kidney

Liver

Intestine



Tandem MS of a quercetin diglucuronide metabolite from rat intestine



Identification of Metabolites of [2-¹⁴C]Quercetin-4'-Glucoside Detected in Rats.

Peak	Rt (mins)	Compound	[M-H] ⁻ (m/z)	Fragment ions (m/z)
1	16.7	Quercetin diglucuronide	653	477 (M ⁻ -GlcUA), 301[Q](M ⁻ -GlcUA-GlcUA)
2	20.6	Unknown		
3	21.6	Methylquercetin diglucuronide	667	491 (M ⁻ -GlcUA), 315 [MQ](M ⁻ -GlcUA-GlcUA)
4	23.7	Quercetin diglucuronide	653	477 (M ⁻ -GlcUA), 301[Q](M ⁻ -GlcUA-GlcUA)
5	24.3	Methylquercetin diglucuronide	667	491 (M ⁻ -GlcUA), 315 [MQ](M ⁻ -GlcUA-GlcUA)
6	25.1	Quercetin diglucuronide	653	477 (M ⁻ -GlcUA), 301[Q](M ⁻ -GlcUA-GlcUA)
7	26.4	Quercetin diglucuronide	653	477 (M ⁻ -GlcUA), 301[Q](M ⁻ -GlcUA-GlcUA)
8	28.3	Quercetin diglucuronide	653	477 (M ⁻ -GlcUA), 301[Q](M ⁻ -GlcUA-GlcUA)
9	29.6	Quercetin-3-glucuronide	477	301 [Q], (M ⁻ -GlcUA)
10	30.3	Quercetin glucuronide sulphate	557	477 (M-SO ₃), 381 (M-GlcUA), 301 [Q](M-SO ₃ -GlcUA)
11	32.6	Methylquercetin glucuronide sulphate	571	491 (M-SO ₃), 315 [MQ](M-SO ₃ -GlcUA)
12	34.5	Quercetin-4'-glucoside	463	301 [Q](M ⁻ -Glc)
13	35.3	Methylquercetin glucuronide	491	315 [MQ](M ⁻ -GlcUA)
14	36.5	Methylquercetin glucuronide	491	315 [MQ](M ⁻ -GlcUA)
15	37.4	Methylquercetin glucuronide	491	315 [MQ](M ⁻ -GlcUA)
16	37.4	Quercetin glucuronide	477	301 [Q](M ⁻ -GlcUA)
17	44.6	Quercetin	301	
18	51.1	Methylquercetin sulphate	395	315 [MQ](M-SO ₃)
19	51.2	Quercetin sulphate	381	301 [Q] (M-SO ₃)

HPLC-RC retention times, negative ion MS-MS fragmentation patterns and identities of metabolites detected in rat tissues 60 min after oral ingestion of [2-¹⁴C]quercetin-4'-glucoside. Peak number refer to peaks in Figure 1. Q – quercetin, MQ – methylquercetin, GlcUA – glucuronyl unit; Glc – glucosyl unit; [M-H]⁻ - molecular ion.

W. Mullen, R. C. Hartley, A. Crozier, *Jouranl of Chromatography A* **2003**, 1007, 21-29. W. Mullen, B. A. Graf, S. T. Caldwell, R. C. Hartley, G. G. Duthie, C. A. Edwards, M. E. J. Lean, and A. Crozier, *J. Agric. Food Chem.* **2002**, 50, 6902-6909.

Chemical Biology

Detect and identify ROS

Locate and quantify ROS

Ameliorate oxidative stress

Track and assess antioxidants



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Alison Hay



welcometrust



Stuart Caldwell



Caroline Quin